PHYLOGENY AND SPECIES DELIMITATION OF NORTH AMERICAN LECCINOID FUNGI

*BEATRIZ ORTIZ-SANTANA US Forest Service, Northern Research Station, Center for Forest Mycology Research, One Gifford Pinchot Dr, Madison, WI 53726 USA MICHAEL KUO Eastern Illinois University, Charleston, IL 61920 USA

Abstract

Leccinoid fungi include ectomycorrhizal members of the Boletales (Basidiomycota, Agaricomycetes) with scabrous to pseudo-scabrous stipes and non-ornamented basidiospores, commonly collected in summer and fall in diverse forest types throughout North America. Previous phylogenetic work on leccinoid fungi has focused primarily on European species; this project centers around North American species. Using collections made by the authors and herbarium material, the initial focus of the research will be the oak-associated leccinoid fungi, which have traditionally been placed in Leccinum, Boletus, and Leccinellum. Sequences from two nuclear ribosomal DNA regions (LSU and ITS) and from one protein-coding gene (TEF1-alpha) have been amplified, sequenced, and analyzed in combination with morphological and ecological data to resolve the continent's major species groups. The working hypothesis for the research, suggested by previous molecular studies, is that ectomycorrhizal host specificity roughly correlates to major clades among the leccinoid fungi, and that species groups can be further delineated with a combination of molecular and morphological characters. Preliminary results of the ongoing research based on LSU and TEF data are presented.

Materials and Methods

A total of 145 LSU and 82 TEF1 sequences were used in the present study; of these 61 LSU and 53 TEF1 were newly generated from herbarium specimens obtained from the Center for Forest Mycology Research (CFMR, Madison, WI), the Stover-Ebinger Herbarium (EIU, Charleston, IL), the New York Botanical Garden (NY, Bronx, NY), the U. S. National Fungus Collections (BPI, Beltsville, MD) and the personal herbarium of Michael Kuo (Charleston, IL), while 82 LSU and 29 TEF1 were retrieved from GenBank. GenBank sequences primarily represent isolates of material from Europe and Asia, while the newly generated sequences are from the United States and Belize.

The 5' end of the LSU region was amplified with primer pair LROR/LR5, and TEF1 was amplified using primer pair EF1-983/EF1-1567R. Sequences were edited with Sequencher 4.8 and aligned using MAFFT v.6. The alignments were manually adjusted using MacClade 4.08.

The phylogenetic relationships among leccinoid species were evaluated with two DNA sequence datasets (LSU and TEF) and maximum likelihood (ML) analyses. ML was run in the RAxML server, v.7.2.8, under a GTR model with 100 rapid bootstrap replicates. The LSU dataset included sequences of about 19 species of *Leccinum/Leccinellum* and at least one species of the *Boletus*, *Chamonixia*, *Harrya*, *Octaviania*, *Roosbeevera*, *Sutorius* and *Xerocomus*, and the TEF dataset included about 15 species of *Leccinum/Leccinellum* and at least one of *Boletus*, *Harrya*, *Octaviania*, *Sutorius* and *Xerocomus*.

Results

In the analysis of the LSU dataset (FIG 1), leccinoid fungi were grouped into four moderately to strongly supported main clades: the **Leccinum (A)**, **Harrya (B)**, **Xerocomus (C)** and **Sutorius (D)** clades.

Within the **Leccinum clade** six groups were obtained: the **Leccinum group** (species of *Leccinum* sections *Leccinum* and *Scabra*), the **Octaviania group** (*Octaviania* species), the **Leccinum viscosum group** (species of *L. viscosum*, *L. violaceotinctum* and *L. quercophillum*), the **Leccinellum group** (species of *Leccinellum* and *Rossbeevera*), the **Boletus longicurvipes group** (species of *Boletus* section *Pseudoleccinum* ss. Smith & Thiers) and the **Chamonixia group** (*Chamonixia caespitosa*); the Leccinum viscosum and Leccinellum groups were not statistically supported.

The **Harrya group** includes *H. chromapes* (previously *Leccinum* section *Roseoscabra*) and *H. atriceps*. The **Xerocomus clade** includes species of *Xerocomus* (section *Xerocomus*), *Boletus hortonii*, *B. rubropunctus* and *Boletus subglabripes* (*Boletus* section *Pseudoleccinum* ss. Smith & Thiers). The **Sutorius clade** includes *S. australiensis* and *S. eximius* (previously *Leccinum* sect. *Eximia*). In the analysis of the TEF dataset (**FIG 2**) these four main clades were also recovered and the Leccinum viscosum and Leccinellum groups were also not statistically supported.

Discussion/Conclusions

This study presents preliminary results from ongoing research into the leccinoid fungi; only about 20 species of the approximately 134 species of *Leccinum/Leccinellum* that have been described were included. Therefore, further work is needed, including additional taxon sampling and multilocus analyses, to determine species delimitations among the leccinoid fungi of North and Central America.

Our results suggest that the genus *Leccinum* needs to be revised. LSU and TEF molecular data indicate that species of *Leccinum* sections *Leccinum*, *Scabra* and *Luteoscabra*, along with *Boletus longicurvipes*, are related, and that their placement within the same genus should be considered; these species also appear closely related to species of *Rossbeevera*, *Octaviania* and *Chamonixia*.

Leccinum subglabripes does not belong to the genus Leccinum, but it is related to the Xerocomus subtomentosus group. Furthermore, Boletus rubropunctus and Boletus hortonii are not closely related to Leccinum species but to the Xerocomus subtomentosus group, as suggested in previous studies.

Our results also suggest the possibility that some GenBank sequences for leccinoid fungi may represent misidentifications.

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Boletus longicurvipes (BOS-659)

Harrya chromapes (MKUO 09150709)

91 Chamonixia_caespitosa_EU669260

4 Xerocomus_subtomentosus_AF139716_DE

erocomus illudens AF139714 USA

— Xerocomus_perplexus_JQ003702_USA

XEROCOMUS

BOLETUS RUBROPUNCTUS

BOLETUS SUBGLABRIPES

Boletus subglabripes (MKUO-08240506)

(section Pseudoleccinum

BOLETUS HORTONII

ss. Smith & Thiers)

SUTORIUS

100 Xerocomus_cf_impolitus_EU522757_CA

Xerocomus depilatus AF139712 DE

77 Xerocomus hortonii AF139713 USA

Leccinum_subglabripes_07230802_USA_IL

ccinum_subglabripes_09010901_USA_IL

cinum_subglabripes_08240506_USA_IL

ccinum_subglabripes_07070702_USA_IL

eccinum_subglabripes_08310702_USA_IL

eccinum subglabripes 08180901 USA IA

- Boletinellus merulioides AY684153

96 — Sutorius_eximius_JQ327010_CR

Sutorius_eximius_AF139684_USA

- Hydnomerulius_pinastri_GU187580

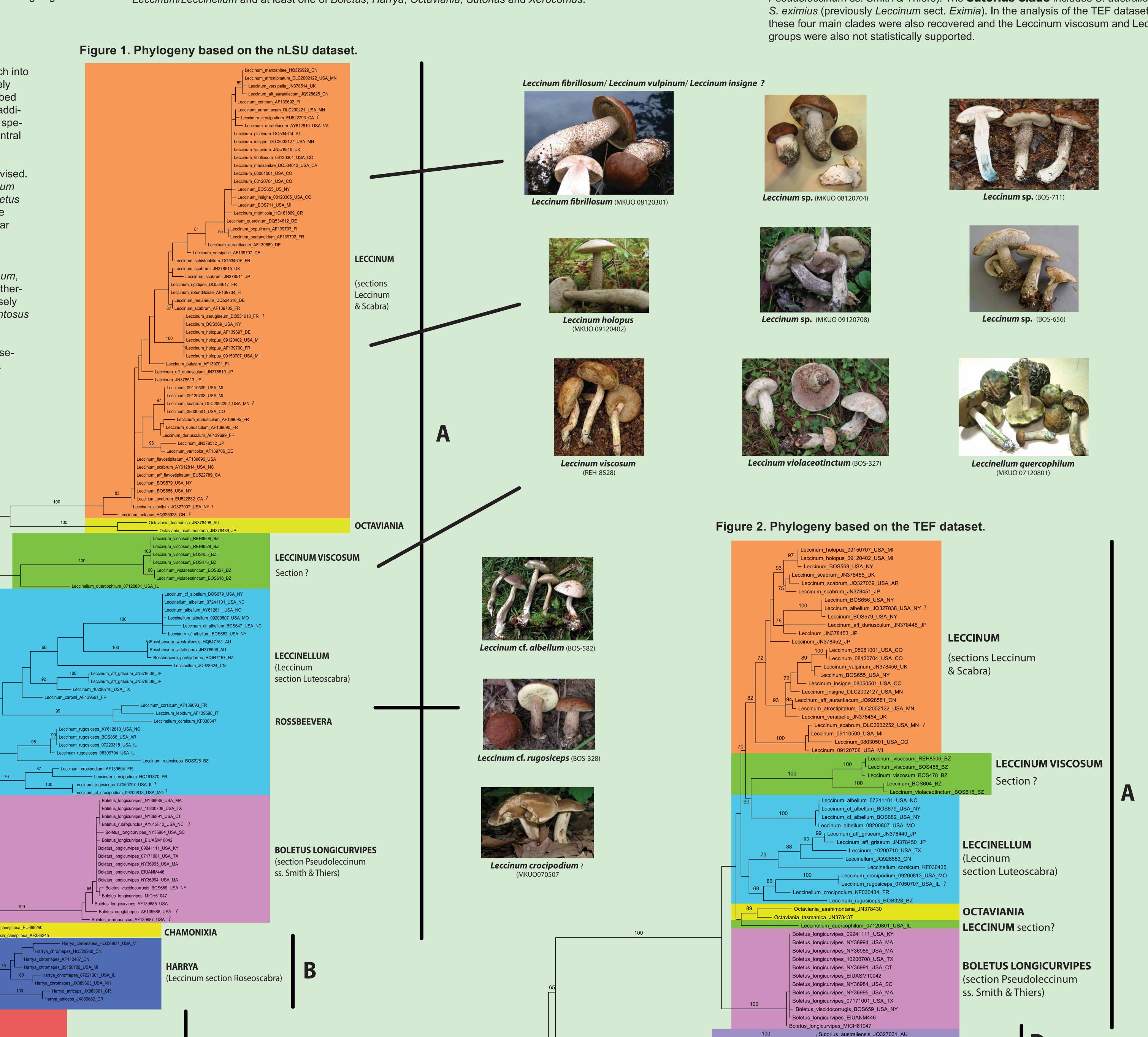
Sutorius_eximius_JQ327004_USA_NY

Sutorius_australiensis_JQ327005_AU

Sutorius australiensis JQ327006 AU

Boletus rubropunctus NY792788 USA NY

- Xerocomus impolitus AF139715 DE



100

100

0.05

———— Sutorius_australiensis_JQ327032_AU

Leccinum_subglabripes_07230802_USA_IL

Leccinum_subglabripes_08160504_USA_IL

Leccinum_subglabripes_08180901_USA_IA

Leccinum_subglabripes_08310702_USA_IL

Leccinum_subglabripes_07070702_USA_IL

Leccinum_subglabripes_08240506_USA_IL

- Boletus_subglabripes_KF030404_USA_NH

— Xerocomus impolitus JQ327034

Leccinum_subglabripes_09010901_USA_IL

— Harrya_chromapes_09150709_USA_MI

73 Harrya_chromapes_KF112270_CN

rrya chromapes 07221001 USA IL

Boletinellus_merulioides_DQ056287Hydnomerulius_pinastri_GU187708

└─ Harrya chromapes JX889703 USA NH

Boletus_rubropunctus_NY1193924_USA_NY

Boletus rubropunctus NY292788 USA NY

Sutorius_eximius_JQ327030_CRSutorius_eximius_JQ327029_USA_NY

SUTORIUS

BOLETUS RUBROPUNCTUS

BOLETUS SUBGLABRIPES

(section Pseudoleccinum

section Roseoscabra)

BOLETUS HORTONII

ss. Smith & Thiers)

XEROCOMUS

HARRYA

(Leccinum